OS114 2002 Ocean Sciences Meeting

the primary food for 4.5 years from hatching to meta-morphosis. Larval shell increased in width and length from 0.24 x 0.20 to 3.0 x 3.4 mm (greatest dimensions perpendicular and parallel to shell axis). Mean shell sizes increased slowly during their later years as lar-vae. The final protoconch width was similar to the 2.7 and 2.8 mm width on shells of field collected snails, in-dicating that size at metamorphosis was close to that transverse spread across the yelar lobes. The total transverse spread across the yelar lobes was as great as 2 cm. Fouling on shells of old larvae included di-atoms and vorticellid and folliculinid ciliates. The lar-vae spent their last two years mostly on the bottom of the gently stirred glass jars with yelar lobes extended. They did not metamorphose until the 11 survivors, out of the 4000 initially cultured, were presented with sub-They did not metamorphose until the 11 survivors, out of the 4000 initially cultured, were presented with sub-tidal rocks and shells as substratum. Five metamor-phosed juveniles survived and grew, the two females first reproducing 3.5 years after metamorphosis. This extraordinarily long larval period in culture may not re-flect the natural duration but indicates the possibility of larval periods far longer than previously suspected. The larval duration exceeded the time from metamor-phosis to first reproduction and greatly exceeded pre-vious estimates of maximum time for larval transport in ocean currents.

OS21G-141 0830h POSTER

Response of Anchovy (Engraulis ringens) off Northern Chile to the 1997-1999 El Niño - La Niña Sequence

Jose L. Blanco¹ (jlblanco@ccpo.odu.edu)

Patricio Barria² (pbarria@ifop.cl)

Jorge Castillo 2 (jcastill@ifop.cl)

Larry P. Atkinson¹ (atkinson@ccpo.odu.edu) ¹Old Dominion University, 768 w 52nd street, Norfolk, VA 23508, United State

²Instituto de Fomento Pesquero, Valparaiso, Chile

 2 Instituto de Fomento Pesquero, Valparaiso, Chile Anchovy are known to be strongly dependent on en-vironmental condition over their life cycle. The 1997-1999 El Niño-La Niña sequence presented an excellent opportunity to observe their response to large scale changes in oceanographic conditions off northern Chile. Observations from a sequence of cruises sponsored by Chilean Fisheries Institute (IFOP) and other data al-lowed us to determine that response. The data include spatial and vertical anchovy distribution off northern Chile (18°S - 24°S), sea surface temperature (SST) im-ages, CTD data from seasonal cruises and wind stress from coastal stations. The data on anchovy came from industrial fisheries catch data and anchovy tertical and spatial distributions from seasonal acoustic cruises. The data was separated into coastal and oceanic for statistical analysis. The sequence stated in March 1997 with warm and salty waters propagating along the South Americas coast. Maximum temperature and salinity anomalies in surface water were 4°C and 0.6 re-septively. The anomaly reached depths of 400 m and was within 50 km of the coast in December 1997. After the maximum in December the anomalies diminished to rapased into a cold phase in which the system is highly pounctive. At the beginning to the El Niño, the anchovy re-

productive

At the beginning to the El Niño, the anchovy re-sponded by concentrating along the coast (first 20 km) where they were more vulnerable commercial fisheries. During this time mortality was high because of the ef-fective fishery (high landings) and environmental stress was high because of El Niño conditions. Later the an-hour mende continueral along the coast and decret in was high because of El Niño conditions. Later the an-chovy moved southward along the coast, and deeper in search of the food and colder water that they prefer (14 to 20° C). They were located between 40 m and 80 m depth (20 to 40 m deeper than normal) near the bottom and far out of range of the net of commercial fishing. They remained at the deeper more southern location until the conditions returned to normal in August 1998. When the anchovy reappeared at their normal locations of fisheries, the biomass was relatively high. This surof fisheries, the biomass was relatively high. This sug-gests that with the onset of upwelling during La Niña period, the stock began their slow recuperation.

OS21G-142 0830h POSTER

Prey Selection, Grazing Rates and Egg Production of Calanoid Copepods in the Firth of Forth, Scotland

 $\frac{\text{Lee Paul Patchell}^1 (+44 (0)131 451 3469;}{1.\text{patchell@hw.ac.uk}}$

- Andrew Garwood Hirst¹ (+44 (0)131 451 3469; a.hirst@hw.ac.uk)
- ¹Department of Biological Sciences, Heriot-Watt University, Edinburgh EH14 4AS, United Kingdom

In this study we investigated rates of fccundity, feeding, and prey selectivity by the dominant calanoid copepods at a fixed station in the Firth of Forth, an

estuarine system on the east coast of Scotland. At fort-nightly intervals over a full annual cycle copepods were collected, and adult females of the dominant species incubated with natural seawater over 24 hours under closely controlled conditions. Species examined in-cluded Acartia bifilosa, A. discaudata, A. clausi, Cen-tropages hamatus and Temora longicornis. Egg pro-duction rates by these copepods and their grazing rates upon nano- and microplankton were quantified by mi-croscope counts. We describe seasonal feeding rates, and prey selection for each of these species. All species cleared ciliates at the highest rates, with the exception of the highly motile species Mesodinium rubrum; while the tintinnids were generally cleared more than the aloricate ciliates. With the exception of Gyrodinium spp., all dinoflagellates were grazed at rates lower than other microplanktonic taxa, possibly indicating active avoidance by copepods. Egg production peaked in May for Acartia discaudata and Temora longicornis with maximum rates of 32 and 33 eggs per female per day re-spectively, rates peaked in mid-July for Acartia clausi and Centropages hamatus at 35 and 70 eggs per female per day respectively. per day respectively.

OS21H HC: 318 A Tuesday 0830h Marine Microbial Genomics Session

Presiding: G Taroncher-Oldenburg, Princeton University; B Ward, Princeton University

OS21H-01 0830h INVITED

The Marine Environment From a Cyanobacterial Perspective

Brian Palenik¹ (858-534-7505; bpalenik@ucsd.edu) Ian Paulsen²

Bianca Brahamsha¹ Rebecca Langlois¹

 $Frank W Larimer^3$

- Scripps Institution of Oceanography, University of California, San Diego, La Jolla, CA 92093-0202, United States
- ² The Institute for Genomic Research, 9712 Medical Center Dr., Rockville, MD 20850, United States
- ³Oak Ridge National Laboratory, 1060 Commerce Park, Rm 211, MS-6480, Oak Ridge, TN 37831-6480, United States

Park, Rm 211, MS-6480, Oak Ridge, TN 37831-6480, United States The genome sequence of the marine cyanobacterium Synechococcus strain WH8102 is nearly completed. This microorganism was chosen because cyanobacte-ria similar to WH8102 are ubiquitous and signifi-cant primary producers in oligotrophic marine envi-ronments. In addition this strain posseses a unique type of prokaryotic motility and is amenable to ge-netic manipulation. The genome is estimated to be 2.7 Mb with approximately 2390 ORFS. The transporter complement of Synechococcus WH8102 was analyzed by screening its genome against a database of known and putative transporters by BLAST and HMM-based analyses. Approximately eighty transport systems were identified comprising 130+ genes. Comparison with the transporter complement of other complete genomes indicated that WH8102 has an emphasis on transport of inorganic anions, in particular with multiple trans-porters for nitrate, sulfate and chloride. In terms of organic nutrients it is predicted to transport systems yof amino acids and a limited number of sugars. The transporters and other activities of the cell are coor-ponent regulatory systems compared to the freshwater cyanobacterium Synechocystis PCC6803. Ultimately the WH8102 genome will provide us with a better un-derstanding of how cyanobacteria perceive and respond to the marine environmet.

OS21H-02 0900h

Defining the Fe Scavenging Mechanism of Marine Synechococcus Using Molecular Approaches

A Webb¹ (ewebb@whoi.edu)

James W Moffett² (jmoffett@whoi.edu)

- John B Waterbury¹ (iwaterbury@whoi.edu)
- ¹Department of Biology, Woods Hole Oceanographic Institution, Woods Hole, MA 02543, United States
 - ²Department of Marine Chemistry and Geochemistry Woods Hole Oceanographic Institution, Wood Hole, MA 02543, United States

Cyanobacteria of the genus Synechococcus are promi-nent components of the marine biosphere that con-tribute significantly to primary productivity. Recent field studies have shown that growth by marine Syne-chococcus can be limited by bioavailable Fe. Despite the demonstrated physiological importance of Fe for open occan Synechococcus, we know very little about how they scavenge it from the environment. Recently the Joint Genome Institute and the Department of Energy se-quenced the complete genome of Synechococcus WHS102. In an effort to understand how these cyanobacteria persist during periods of Fe deprivation, we have per-formed physiological and genomic experiments to depersist during periods of Fe deprivation, we have per-formed physiological and genomic experiments to de-fine their Fe scavenging mechanism. By determining how these microorganisms respond to Fe stress, we will gain insight into how and when this important trace el-ement can limit their growth in situ. This knowledge will greatly increase our understanding of how marine Fe cycling impacts oceanic processes.

OS21H-03 0915h

A Search for Cobalt Proteins in Synechococcus: Overexpression of the Gamma Carbonic Anhydrase Enzyme and a Genome Wide Analysis of Cobalt Limited Cells

Mak A. Saito¹ (609-258-2612; msaito@princeton.edu)

Todd W. $Lane^1$ (tlane@princeton.edu)

Gaspar Taroncher-Oldenburg¹ (gto@princeton.edu)

Francis M.M. Morel¹ (morel@princeton.edu)

Bess B Ward¹ (bbw@princeton.edu)

¹Department of Geosciences, Princeton University, Princeton, NJ 08544, United States

Princeton, NJ 08544, United States The marine cyanobacteria have an absolute cobalt requirement for which zinc cannot substitute. Cobalt concentrations in seawater are extremely low and may therefore affect primary productivity or phytoplankton species abundance. Yet it is not known what the pre-dominant biochemical function of cobalt is in the sur-face oceans. We hypothesize that the metal in the ac-tive site of the gamma carbonic anhydrase of marine cyanobacteria may be cobalt and this may be a ma-jor use of cobalt in the surface oceans. The gamma carbonic DNA of marine Synechococcus WH8102, cloned, and the correct sequence was verified by complete dounomic DNA of marine Synchococcus WH8102, cloned, and the correct sequence was verified by complete dou-ble strand sequencing. The gene was then cloned into an expression vector, overexpressed into *E. coli*, and pu-rified and characterized from inclusion bodies. Strate-gies for refolding the protein with cobalt and zinc will be presented. Results from a parallel whole genome mi-croarray study of cobalt limited Synchococcus WH8102 to identify other potential cobalt dependant proteins and pathways will be discussed in the context of our limited understanding of the biochemical function of cobalt in the cyanobacteria.

OS21H-04 0930h INVITED

Chronology of the Transcriptional Response to Environmental Stimuli in Cyanobacteria

Gaspar Taroncher-Oldenburg¹ (gto@princeton.edu); Mak A. Saito² (msaito@princeton.edu); Ryan T.

Gill³ (rtg@colorado.edu); Gregory Stephanopoulos⁴ (gregstep@mit.edu); Francois M.

Morel² (morel@princeton.edu); Bess B. Ward² (bbw@princeton.edu)

¹Princeton Environmental Institute, Princeton University, Princeton, NJ 08544, United States

²Geosciences Department, Princeton University, Princeton, NJ 08544

- ³Department of Chemical Engineering, University of Colorado, Boulder, CO 80309, United States
- ⁴Department of Chemical Engineering, Massachusetts Institute of Technology, Cambridge, MA 02139, United States

United States Aquatic microorganisms exhibit a wide range of adaptation mechanisms to a variety of changes in their environment. The response to a new stimulus can oc-cur at different levels, but is most often centered on fine tuned variations in the transcriptional profile of the cell. In this contribution we will illustrate the molecular chronology of such changes as they occur at the whole genome level. Two different examples of re-sponse to environmental stimuli in cyanobacteria are presented. First, we will analyze the transcriptional program elicited by the light to dark transition in the freshwater cyanobacterium Synechozystis sp. PCC6803. Special attention is given to genes related to light harvesting, energy transformations and central carbon metabolism. In a second example, the whole genome re-sponse of the marine cyanobacterium Synechozoccus sp.

Cite abstracts as: Eos. Trans. AGU, 83(4), Ocean Sciences Meet. Suppl., Abstract #######, 2002.

WH8102 to changes in levels of the trace metal cobalt is WH8102 to changes in levels of the trace metal cobalt is investigated. Common characteristics to the transcrip-tional profile dynamics in both cases are coregulation of polycistronic genes and overshoot dynamics that are ubiquitous in most biological systems. Results on spe-cific genes and pathways will also be presented.

OS21H-05 1030h INVITED

Creating Large and Small Insert Chromosomal Libraries From Naturally Occurring Microbial Populations: Nuts And Bolts

Christina M Preston¹ (831-775-1754; preston@mbari.org)

Oded Beja^{2,3} (beja@techunix.technion.ac.il)

Grieg F Steward^{2,3} (gsteward@cats.ucsc.edu)

Edward F Delong¹ (delong@mbari.org)

- ¹Monterey Bay Aquarium Research Insitute, 7700 Sandholdt Rd , Moss Landing, CA 93950, United States
- $^2\,\mathrm{Department}$ of Biology, Technion, Israel Institute of Technology, Haifa 32000, Israel
- 3 Ocean Sciences Department, E&MS A44 University of California 1156 High Street, Santa Cruz, CA 95064, United States

Secrets of the natural microbial world, including the specific biological properties and function of mi-crobes in Nature, can in part be revealed by directly sequencing the genomes of these organisms. There are sequencing the genomes of these organisms. There are a number of different strategies and approaches for archiving and extracting the genomic sequence from mi-crobes present in natural populations. One approach, of course, is simply to cultivate the microbe, and se-quence its genome using now standard shotgun se-quencing strategies. This approach is not always prac-tical, feasible, or possible. Another approach is to construct large or small insert genomic libraries from mixed microbial assemblages, and either randomly, or systematically, sequence and analyze microbial genomic fragments. The different strategies and vectors in-clude small insert shotgun libraries (insert size around 3 kilobases (kb)), fosmid or cosmid libraries (insert sizes around 40kb), and bacterial artificial chromosome (BAC) libraries (inserts up to 200 kb), and each apsizes around 40kb), and bacterial artificial chromosome (BAC) libraries (inserts up to 200 kb), and each ap-proach has its strengths and weaknesses. We have had success with preparing libraries from naturally occur-ring bacterial or viral populations. Libraries currently being analyzed include those from bacterial and viral populations of marine plankton, and microbial commu-nities in anaerobic marine sediments. These are now providing significant new insight into the genomic and functional properties of microbes from diverse ecosys-tems

OS21H-06 1100h

A General Method for Growing Unculturable Microorganisms

Tammi Kaeberlein¹ (781-581-7370;

eber@ccs.neu.edu)

Kim Lewis¹ (617-373-8238; k.lewis@neu.edu)

Slava Epstein¹ (617-373-4048; s.epstein@neu.edu)

¹Department of Biology, Northeastern University, Boston, MA 02115

The majority of microorganisms from the environ-ment resist cultivation in the laboratory. Several Bac-terial and Archaeal groups at the division level have been identified with no known cultivable representa-sights into the spectacular diversity of uncultivated mi-croorganisms. However, it does not provide access to the actual cells and means to grow them. Such access would be highly desirable for both basic and applied environmental sciences. Here we report a new gen-eral method for growing previously uncultivated mi-croorganisms. Diffusion growth of these microorgan-isms in pure culture by providing a simulated nat-nal environment. We will discuss design of the cham-bers, provide examples of first uncultivables growing in our laboratory, characterize these novel organisms us-ing 16S rRNA data, and outline the potentials of the new method. The majority of microorganisms from the environ-

OS21H-07 1115h INVITED

Nitrogen Assimilation: from Genomes to Gene Expression in the Oceans

Jonathan P Zehr (831-459-4009; zehrj@cats.ucsc.edu)

Dept. of Ocean Sciences, University of California. Santa Cruz, CA 95064, United State

Nitrogen (N) assimilation is intimately linked to controls on primary production in the open ocean. The application of the polymerase chain reaction has pro-vided information on the diversity and distribution of the genetic information for specific N utilization path-ways, from transport to assimilatory nitrate reductase (nar and nas) genes indicates that the genetic capa-bility for different N metabolic pathways is dispersed throughout microbial phyla, and that genotypic as well as phenotypic differences among microbial assemblages are likely to be important in determining the pathways of nitrogen use, and the ecological distribution and ac-tivities of individual species. Gene expression assays provide ways of identifying important players in nitro-Nitrogen (N) assimilation is intimately linked to provide ways of identifying important players in nitro-

Invites of manual processors of the provide ways of identifying important players in nitro-gen metabolism. Assimilatory nitrate reductase genes are less widely distributed among cyanobacteria and heterotrophic bacteria than might be expected. It has recently been shown that Prochlorococcus does not contain ni-trate reductase genes, which suggests that the genome has eliminated unnecessary genetic information for or-ganisms living in oligotrophic surface ocean waters. Nitrogenase-containing microorganisms are more di-verse and abundant in the open ocean than previously believed, but relatively few major clades have been discovered. There is a high diversity of nitrogenase-containing microorganisms in some high fixed N envi-ronments such as the Chesapeake Bay and deep wa-ters of Mono Lake. Thus, ecosystem comparisons sug-gest that nitrogenase diversity is not selected for by

ronments such as the Chesapeake Bay and deep wa-ters of Mono Lake. Thus, ecosystem comparisons sug-gest that nitrogenase diversity is not selected for by N-limiting conditions, and that genes are not necessar-ily eliminated from the genome in non-N-limiting en-vironments. The data indicate that there are multiple ecological and evolutionary forces selecting for genome composition of individual species or phylotypes, includ-ing selection at the organism or community level. Over 50 microbial genomes have been sequenced or are in the process of being sequenced, as well as ge-nomic fragments from marine environments. Statis-tics from these prokaryotic genomes indicate that 1) N metabolism genes are not well-represented in these genomes or that the genes have not been annotated cor-rectly, 2) that the genomes that have been sequenced may not be good models for the study of N assimila-tion in the environment and/or 3) that nitrate might not be used by many prokaryotic microorganisms. Ge-nomic information provides important contextual in-formation, including gene arrangements and new genes to target in environmental studies, yet one of the ma-jor limitations of genomic information is that some biogeochemically-important microorganisms can be rel-atively low in abundance, making it difficult to obtain atively low in abundance, making it difficult to obtain biogeochemically-relevant information directly from ge-nomic fragments obtained from the environment.

OS21H-08 1145h

A Multisystematic Approach Towards Understanding the Metagenome of the Episymbiotic Community Associated With Alvinella pompejana

Barbara J. Campbell¹ (bjc@udel.edu); Maureen Dolan² (maureen.dolan@USA.dupont.com); Kathryn J. Coyne¹ (kcoyne@udel.edu); Benjamin

Wheeler¹ (jaywh@udel.edu); Jens J. Hyldig-Nielsen³ (jhyldig@bostonprobes.com); Robert A. Feldman⁴ (robert.feldman@am.apbiotech.com); Jeffrey

Stein⁵ (jstein@quorex.com); S. Craig Cary¹

- (caryc@udel.edu)
- ¹University of Delaware, College of Marine Studies, 700 Pilottown Rd., Lewes, DE 19958
- ²Dupont Agricultural Biotechnology, Delaware Technology Park, Suite 100, PO Box 6107, Newark, DE 19714
- 3 Boston Probes, Inc., 75E Wiggins Ave., Bedford, MA 01730
- $^4\,\mathrm{Molecular}$ Dynamics, Inc., 928 E. Arques Ave., Sunnyvale, CA 94086
- ⁵Quorex Pharmaceuticals, Inc., 2075-J Corte Del No-gal, Carlsbad, CA 92009

⁶ Guorex Pharmaceuticals, Inc., 2075-J Corte Del No-gal, Carlsbad, CA 92009 A diverse episymbiotic bacterial community is asso-ciated with the tube-dwelling polychaete, Alvinella pom-pejana. This association exists in an extreme deep-sea hydrothermal vent biotope characterized by high con-centrations of heavy metals and the steepest thermal gradient experienced by any organism yet described. Detailed rRNA analysis of the episymbiotic commu-nities associated with A. pompejana demonstrates the dominance of a diverse assemblage of a single subdi-vision (epsilon Proteobacteria). Because of the complex nature of this association, no specific roles have been defined for this unique symbiosis by habitat charac-terizations, in situ enzyme assays, classical cultivation techniques or molecular analysis. Initial approaches to investigate the symbionts centered on rRNA analysis, where two filamentous epsilon Proteobacteria dominate the microbial community. Current work in our labo-ratories centers on a whole genomics approach. High throughput sequencing efforts allow the creation of a metagenome, where genomes of a diverse ecological unit

2002 Ocean Sciences Meeting OS115

are cloned and sequenced. These collective genomes can be thought of as a core genome, containing all the genes necessary to perform the diverse biogeochemical reactions that make up a specific community function. Initial analyses presented here include the sequencing of two large insert fragments (approximately 40kb each) specifically linked to the two dominant symbiont phy-lotypes. We have also begun to use a metagenome ap-proach in order to understand the symbiont commu-nity at a functional ecological level. A cDNA library has been made from ribosomal depleted RNAs isolated from an A. pompejana symbiont community from $g^0 N$, East Pacific Rise. Preliminary sequence analysis from this library will also be presented.

OS21I HC: 314 Tuesday 0830h **Reforming Education in the Ocean** Sciences for All Citizens II

residing: J Cherrier, Florida A&M UniversityUniversity; C Thoroughgood, University of

Delaware; P Coble, University of South Florida

OS21I-01 0830h INVITED

The Art of the Possible, The Science of the Priorities: Educational Opportunities for the Ocean Sciences

F

Arthur R.M. Nowell (206-543-7160; nowell@ocean.washington.edu)

College of Ocean and Fishery Sciences, University of Washington, Box 355350, Seattle, WA 98195, United States

United States Graduate education in our field is changing as the focus of research moves to address the questions driven by the concerns and values of the twenty-first cen-tury. Undergraduate education is changing with the shift from teaching to learning, especially with the shift to experiential learning and collaborative learning. In ocean sciences we are coming to terms with how to in-tegrate collaborative learning and experiential learning into our mandates for teaching and research on cam-pus. The vast responsibility of public involvement and education still is open. At the K-12 levels four broad terms dominate the agenda, namely standards, assess-ment, technology and communication. How can ocean sciences help teachers gain the knowledge of the con-tent areas and the skills of science for elementary level teaching? How can ocean sciences help teachers de-velop among their students problem solving skills? How can ocean sciences help students become adept at us-ing technology in problem solving? And lastly, how can ocean sciences help students select and discuss a problem and its potential solutions and thereby demon-strate problem solving skills? I will discuss the range of possibilities and suggest some ideas that involve not just ourselves at research focused institutions, but also involve agencies, their staff and the chances for a higher level of collaboration. Graduate education in our field is changing as the level of collaboration.

OS211-02 0900h

Inquiry Based Learning and Assessment in General Education Science Courses

 $\underline{\operatorname{Bob}\ \operatorname{Chen}}^1\ ({\tt bob.chen}@{\tt umb.edu})$

Daniel Brabander¹ (daniel.brabander@umb.edu)

¹U. Mass. Boston, ECOS 100 Morrissey Blvd, Boston, MA 02125, United States

MA 02125, United States Two major goals of General Education Science Courses are to recruit top students into science and to provide a large number of non-science majors with the critical thinking skills that they may use throughout their adult lives. In this way ocean and/or environmen-tal sciences may become an integral part of the daily lives of the general public. While most educators agree that effective science learning occurs in small groups with hands-on experiential learning and interaction be-tween the teacher and student, many administrators prefer to maximize university resources by increasing class sizes. sizes

In an attempt to provide quality science instruction In an attempt to provide quality science instruction to a large General Education audience, Introduction to Environmental Sciences was offered to 230 students at UMassBoston in the Fall of 2001. Due to unique cir-cumstances, 50 of the students were required to attend weekly discussions (ES 120; 10-15 students/section) while the other 180 were required to write 4 papers (ES 101). All 230 students attended the same two 75 minute lectures each week. In class peer-instruction, group activities, and a class web site were used to en-gage students. On a traditional exam of multiple choice and short answer questions, ES 120 students performed

Cite abstracts as: Eos. Trans. AGU, 83(4), Ocean Sciences Meet. Suppl., Abstract ########, 2002.